Structure-Activity Relationship of Aldose Reductase Inhibitors Based on X-Ray Crystal Structures of Oxazolecarbamate Derivatives

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SUMMARY

In order to elucidate the key atoms and/or stereostructures necessary for the inhibitory emergence of aldose reductase, crystal structure determinations were carried out for 11 oxazolecarbamate analogues, which have similar chemical and physicochemical properties but different inhibitory activities. The molecular conformations, revealed by X-ray analyses, were also ascertained to be energetically stable from theoretical confor-

mational energy calculations. A surprising degree of conformational similarity was observed for the potent inhibitors. The analyses of the quantitative structure—activity relationships showed that the molecular conformation and the dipole moment, as well as the hydrophobicity at the oxazole C5-site, were important for high activity.

Aldose reductase (EC 1.1.1.21), one of the NADPH-dependent aldoketo reductases, is the first enzyme of the polyol pathway and catalyses the reduction of hexoses to their respective sugar alcohols. Sorbitol accumulates in the lens (1, 2), peripheral nerves (3, 4), and retina (5, 6) of diabetic humans and animals. Elevated sorbitol levels result in a loss of osmotic integrity and subsequent cellular damage (7, 8), which are linked to development of some complications of diabetes such as cataracts, neuropathy, retinopathy, and nephropathy (9, 10). Because excessive sorbitol is produced from glucose by aldose reductase, increased importance has been placed on the development of potent aldose reductase inhibitors, which are viewed as a means to reduce chronic diabetic complications. Several compounds with different chemical structures, such as sorbinil (11), ONO-2235 (12), and tolrestat (13), were suspected to be therapeutic. Recently, benzyl 4-isopropyl-5-phenyl-2-oxazolecarbamate was found to be a potent aldose reductase inhibitor

In order to develop more potent inhibitors, it is important to elucidate the key atoms, and their spatial orientations, necessary for activity emergence (16). A possible methodology is to compare the stereostructures in a series of compounds with similar chemical and physicochemical properties but different pharmaceutical activities. Because the inhibition for aldose reductase in rabbit lens has already been elucidated for the 2-oxazolecarbamate analogues (14, 15) listed in Table 1, we have undertaken in this study the X-ray single crystal structural

analyses of these compounds (except 4) and have considered the structure—activity relationship.

Materials and Methods

All compounds have been chemically synthesized (14, 15).

X-ray analyses. Various attempts to crystallize compound 4 were unsuccessful. The crystallographic data for the other compounds are given in Table 2. Unit-cell dimensions and diffraction intensities were measured in a similar manner with graphite-monochromated Cu Ka radiation ($\lambda = 1.5418 \text{ Å}$) on a Rigaku AFC-5 computer-controlled diffractometer. The unit-cell parameters were determined by a least squares fit to 2θ angles for 20 to 25 reflections (30° < 2θ < 60°). The crystal density was measured by the flotation method using a CCL/ C_6H_6 mixture. The collected intensity data with $\omega - 2\theta$ scan technique were corrected for Lorentz and polarization effects but not for absorption. Non-hydrogen atom positions were located by a combination of direct method (17) and Fourier syntheses and refined by a blockdiagonal least squares method using anisotropic thermal parameters. The positions of the geometrically ideal hydrogen atoms were calculated, which were verified on respective difference Fourier maps, except for compounds 9 and 12, and included in subsequent refinements (only in the calculations of structure amplitudes for compounds 9 and 12) with isotropic temperature factors. The function minimized was Σ $w(|F_o| - |F_c|)^2$, where $|F_o|$ and $|F_c|$ are the observed and calculated structure amplitudes, respectively. The weighting scheme used for the refinement was the following: w = a for $F_o = 0.0$, and $w = 1.0/(\sigma(F_o)^2)$ $+ b |F_o| + c |F_o|^2$ for $F_o > 0.0$, where $\sigma(F_o)^2$ is the standard deviation of the intensity based on counting statistics. The values of a, b, and c

TABLE 1

Chemical structures used for study and their inhibitory activities, as well as atomic numbering schem

Compound	R′	К"	IC ₅₀
		R' 4 N3 27 28 5 21 22 24 NHCOOCH ₂ 26 29 23 25 27' 28'	μм
1	42, 43, 41 (CH₃)₂CH	53 52 54 53' 52'	0.35
2	CH₃CH₂	33 52	0.84
3	CH₃		2.45
4	(CH₃)₃C		12.9
5	н		15.6
6	н	52, 53, 54, 51 (CH₃)₃C	89.5
7 9	CH₃ H	CH₃ CH₃	418 770
8		NHCOOCH₂CH₃	>500
	R' O	22 23 21 24 22' 23'	
10	41 CH₃	58 57 55 53 51 56 CH₂OOCNH 56 54 52	4010
11	48 47 45 43 41 46 CH ₂ OOCNH 48' 47'	н	œ
12	4 3 5	2_NHCOOCH2	œ

in the final refinement are also listed in Table 2, as well as the discrepancy indices, $R=(\sum ||F_o|-|F_c||/\sum |F_o|)$ and $R_w=[\sum w(|F_o|-|F_c|)^2/\sum w|F_o|^2]^{1/2}$, and goodness of fit, $S=[\sum w(|F_o|-|F_c|)^2/(M-N)]^{1/2}$, where M and N are the numbers of reflections used for refinement and atomic variables, respectively. For all crystallographic computations, the UNICs programs (18) were used, and the

atomic scattering factors were cited from *International Tables for X-ray Crystallography* (19). The crystallographic details will be published elsewhere. Bond lengths and angles are given in Table 3. The atomic numbering used for all compounds is shown in Table 1.

Conformational energy calculation. The energy for conformational analysis was calculated by the molecular mechanics method



MOLECULAR PHARMACOLOGY

Summary of crystallographic data and structure refinement procedures

Solvent for crys-	1 S Ethanol/ethylacetate Benzene/n-hexane Chloroform/n	2 Benzene/n-hexane	3 Chloroform/n-	5 Chloroform/n-	6 Methanol/water	8 7 8 Methanol/water Methanol/water Chloroform/n-	8 Chloroform/n-	9 Methanol/water	9 12 Methanol/water Methanol/water Methanol/water	11 Methanol/water	12 Methanol/water
Crystal system Space group	Tricki P1	Monodi P2,/n	exane Monocii <i>P2.\n</i>	exane Monocli P2,/c	Monocii C2/c	Tricii PT	Orthorhom Pbcn	Monodi P2,/c	Tricki PT	Monodi P2,/8	Orthorhom Pca2,
Cell constant	į (6	į į	17,000	60 07	44 000 44	377	1000	9,00,11	60 07	0, 110, 01
€	19.469 (/)	9.131 (/)	9.200 (/)	0.928 (1)	19.30 (2)	(4) 000 (4)	20.44 (1)	(0) 500.4	(c) 88.40 (c) 64.60	(2) / (2)	10.045 (Z)
٥, ٩	11.270(3)	18.81	8.785(5)	(9) 927.71	5.458 (4)	8.544 (5)	12.206 (8)	10.48 (2)	9.16 (4)	9.745(2)	(r) 811.6
د' ۷	8.667 (3)	9.680 (8)	19.800 (9)	25.406 (9)	26.22 (3)	7.406 (4)	7.206 (4)	23.28 (6)	6.00 (2)	13.923 (3)	30.78 (1)
α, deg	95.48 (3)	8	8	8	8	110.51 (4)	8	8	107.8 (3)	8	8
8	99.61 (3)	101.07 (3)	99.60	92.90 (4)	109.81 (9)	103.49 (3)	8	90.9 (1)	92.6 (5)	113.65 (1)	8
٨	104.96 (3)	06	8	8	8	94.35 (4)	8	8	96.2 (4)	8	8
Volume, ų	1791 (1)	1631 (2)	1580 (2)	3033 (2)	3087 (5)	929.2 (7)	2326 (2)	1137 (4)	776 (5)	1493.6 (5)	1581.0 (8)
7	4	4	4	œ	80	8	œ	4	8	4	4
Formula/asymetric unit	o unit										
	2 (C.HN.O.)	C.N.H.C	C.H.	2C.H.N.O.	C.H.D.O.	C.H.N.O.	C.H.NO	C.H.,N.O	C.H.	CH.,N.O.	C.HNO.
Om a.cm ⁻³	1.266(3)	1.310 (3)	1.299 (3)	1 270 (2)	1.201 (5)	1.299 (2)	1.321 (4)	1,329 (5)	1.319 (4)	1.318 (6)	1.280 (4)
Dx a.cm-3	1.247	1.313		1 289	1.180	1.300	1.326	1.357	1.319	1.309	1.274
"(Cir Ka) cm ⁻¹	5.93	6.92	7.72	9	6.44	7.36	7.66	7.84	7.05	7.10	6.19
F(0.00)	712	990	648	1232	1168	260	976	488	324	616	640
Scan speed (in		6	4	<u>س</u>	4	4	~	က	8	က	~
20) dec.min-1	1)							
Scan width in A +											
B tan θ											
A, deg	0.80	1.10	1.00	1.10	1.00	1.00	1.00	1.50	96.1	1.20	1.40
B, deg	0.15	0.15	0.15	0.15	0.15	0.15	0.15	0.15	0.15	0.15	0.15
No. of obsd. re-	6126	2772	2685	5168	2619	2132	1999	1956	2304	2548	1363
Hection	90.50	04.50	6	9900	0640	2400	9550	7660	7000	0640	920
No. Used for re- finement	9719	0#67	7617	9999	6103	2012	8	8	500	ot or	8
() ≥ nσ ()	0	7	8	က	0	0	8	က	0	0	8
Coefficient used											
	1.110	0.0	0.0	0.0	0.193	4.731	0.0	0.0	0.384	1.988	0.0
۰	-0.623	-0.170	-0.380	-0.294	-0.195	-0.076	-5.006	0.397	0.0- 440.0-	-0.433	0.380
v	0.032	0.013	0.022	0.022	0.010	0.010	0.224	9000	0.005	0.026	-0.003
	0.075	0.086	0.077	960.0	0.082	0.081	0.104	0.141	0.105	0.081	0.123
ď	0.082	0.127	0.074	0.152	0.097	0.141	0.086	0.201	0.095	0.107	0.071
S	2.644	3.429	2.210	2.091	1.363	2.581	3.261	0.930	1.367	2.125	1.121
2	612	290	273	510	254	220	203	155	273	248	508

Aspet MOLECULAR PHARMACOLOGY

# #	.354 (5) 1.359 (4) .383 (5) 1.377 (5)	1 284	(6)	.384 (5) 1.378 (4) .324 (6) 1.343 (5)	(6) 1.393	(5)	.388 (6) 1.376 (5) .399 (7) 1.397 (7)	(7) 1.403	.374 (8) 1.391 (7) 357 (8) 1.350 (6)						1.347 (4)	1.204 (4)	1.455 (5) 1.502 (5)	1.370 (5) 1.376 (5)	1.370 (7) 1.434 (7) 1.346 (7)	1.379 (8) .400 (5)		.203 (5)	(c) 815.	468 (6) 504 (7)	.378 (7) .381 (7)	.397 (7) .369 (8)
	1.333 (8) 1.3 1.398 (9) 1.3	_		1.39(1) 1.3	_	1.48 (1) 1.3		_	<u>-</u> -	E	1.37(1)	1.40 (1) 1.41 (1)	1.40 (1)	1.34 (1)	(1) 04:-					1.4		7.	<u>-</u>	7. 4.		
•	1.359 (4)	1 204 (4)	1.362 (4)	1.392 (4)		1.449 (4)	1.209 (4)	(1)	1.543 (4)	1.443 (5)	(0) †									1.401 (4) 1.390 (4)		1.393 (5) 1.393 (5)	1.389 (5)	(c) 9/c·1		
7	1.300 (5)	1 222 (4)	1.365 (5)	1.399 (5)	1.481 (6)	1.481 (6)	1.189 (5)	4 946 (5)	1.340 (5)	1.464 (5)	1.376 (6)	1.386 (6)	1.376 (6)	1.383 (7)	(1) toc:				-							
9	1.330 (3)	1 296 (2)	1.370 (3)	1.407 (4)	(£)	1.492 (4)	1 186 (3)	(2) (3)	(S) /SC.1	1.448 (4)	1.374 (4)	1.381 (4)	1.389 (5)	1.373 (5)	(0) 0/8:1					.537	1.533 (6) 1.507 (6)					
8	1.351 (6)	4 276 (2)	1.365 (7)	1.402 (7)	(6)	1.457 (8)	1 209 (7)	(2) (2)	1.335 (/)	1.473 (8)	1.37 (1)	1.41 (E) (S)	1.37 (1)	1.39 (1)	(E)					1.387 (9) 1.392 (9)		1.41 (1)	1.35 (1)	y.		
3	1.343 (6)			1.387 (7)		1.446 (8)	1.185 (7)		1.359 (/)	1.448 (8)	1.369 (9)	1.375 (9)	1.37 (1)	1.39 (1)	(E) 86.1					1.395 (9) 1.388 (9)		1.39 (1) 1.39 (1)	1.41 (1)	1.37 (1)		
	1.335 (3)		1.289 (3) 1.386 (3)	1.410 (3)	1.491 (4)	1.460 (4)	1 194 (4)	(F) to 0.	1.338 (3)	1.459 (3)	1.391 (4)	1.379 (4)	1.412 (5)	1.338 (6)	1.3/0 (6)					1.385 (4) 1.385 (4)		1.383 (5) 1.383 (5)	1.383 (5)	(c) 995.1		
2	1.335 (4)		1.370 (4)	1.414 (5)	1.616 (7)	1.456 (5)	1.377 (4)	(±) 103.1	1.343 (4)	1.465 (4)	1.388 (5)	1.383 (5)	1.386 (6)	1.374 (6)	1.406 (8)					1.395 (5) 1.394 (6)		1.373 (6) 1.397 (7)	1.398 (6)	(/) L96.1		
10	1.347 (3)	(6) (6)	1.374 (3)	1.399 (3)	1.501 (4)	1.456 (3)	1.300 (3)	(6) 261.1	1.337 (3)	1.455 (4)	1.383 (4)	1.375 (4)	1.404 (5)	1.376 (5)	1.536 (4)	(*) too:				1.388 (4)		1.387 (4) 1.382 (4)	1.382 (4)	1.383 (4)		
=	1.338 (3)	(2)	1.287 (3) 1.364 (3)	1.410 (3)	1.496 (4)	1.458 (4)	1.379 (4)	1.132 (4)	1.331 (3)	1.459 (4)	1.300 (4) 1.382 (4)	1.375 (4)	1.373 (5)	1.365 (6)	1.326 (6) 1.526 (5) 1.526 (5)	(6) 186:1				1.395 (4) 1.394 (5)	•	1.387 (4) 1.396 (5)	1.367 (5)	1.384 (5)		



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2222 2228 2222 2228		\$ \$ 2 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5	120 (2) 120 (2) 120 (2)	130 (2) 120 (2) 121 (2) 121 (2)
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		221 21 22 22 22 23 24 24 25 25 25 25 25 25 25 25 25 25 25 25 25	(e) c.o.y.	
115.8 (6) 119.9 (6) 124.2 (6) 103.0 (6) 111.1 (7) 105.9 (6) 137.1 (7) 116.9 (6)	126.5 (6) 126.2 (7) 108.4 (6) 125.4 (7) 115.0 (6)	108.3 (6) 177.1 (7) 172.2 (7) 177.1 (7) 177.1 (7) 177.1 (7) 177.1 (7) 179.2 (8) (8) (9) (9) (9) (9) (9) (9) (9) (9) (9) (9		
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87.81.807.1.7.9 66.66.66.66.66.66.66.66.66.66.66.66.66.	3 (3) (3) (3) (3) (3) (3) (3) (3) (3) (3	110.9 (2) 112.5 (3) 112.5 (3) 112.5 (3) 112.5 (3) 112.5 (3) 112.5 (3) 113.5	119.5 (3) 121. 122.0 (3) 120. 118.5 (5) 118.	66 66 66 66 66 66 66 66 66 66 66 66 66
	6-5 21-22 21-22 22-23 22-24 108 22-24 125-24 125-23 23-24 23-24 125-23 125-23 125-23		48 49 48 5-51-52 52-51-52 115 5-51-53 116 52-51-53 52-51-53	52-53 120 52'-53 119 53-54 121, -53'-54 121, 54-53' 118.
ナ	+448448488	8488688868444444444444	\$ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~	344888

[CHEMLAB-II version (20)]. The energy functions included in the calculation were steric (Lennard-Jones potential), electrostatic (Coulombic), and torsional energies:

$$E_{\text{steric}} = \sum_{i} \sum_{j} (-A_{ij} r_{iij}^{-6} + B_{ij} r_{ij}^{-12})$$
 (1)

$$E_{\text{elec}} = \sum_{i} \sum_{j} 332.0 \cdot Q_{i} \cdot Q_{j} \cdot r_{ij} \epsilon^{-1}$$
 (2)

$$E_{\text{tor}} = A[B + C \cos^{M}(N\theta - \phi)] + D[1 - \cos(E\theta - \psi)]$$
 (3)

where r_{ij} in Eqs. 1 and 2 is the distance (in angstroms) between atoms i and j. The atomic net charge on atom i (Q_i) in Eq. 2 was calculated by the CNDO/2 method (21), and the dielectric constant (ϵ) was taken as 4.0, close to the experimental values for biomolecules in polar media. The A, B, C, D, E, M, N, θ , ϕ , and ψ in Eq. 3 are constants associated with a particular type of bond (22). All calculations of Eqs. 1–3 were performed with the supplied data set (20). The notation of variable torsion angles is shown in Table 4. Among the torsion angles, ω_3 (C2 – N21-C22-024) was set to 180° (trans) for all calculations, because the amide bond shows a strong preference for that value. In the conformational analysis, respective torsions were rotated simultaneously in 10° increments from 0° to 360° (for ω_1 and ω_6 , 0° to 180°), where the starting, eclipsed conformation was 0°.

Dipole moments. The dipole moments $(\mu_x, \mu_y, \text{ and } \mu_z)$ for each molecular conformation were calculated by the CNDO/2 method by using the atomic coordinates obtained by the present X-ray analyses.

¹H NMR measurements. ¹H NMR spectra were measured on a Varian XL-300 (300 MHz for proton) spectrometer equipped with the temperature-controlled accessory (accuracy to $\pm 1^{\circ}$). Chemical shifts were measured with tetramethylsilane as internal standard. The sample concentrations were determined by dry weight and then adjusted to the desired values by dilution. Spectra were recorded at each temperature of 30°, 40°, 50°, 60°, and 70°.

Structure—activity correlation analysis. In developing the correlation equations, the QSAR program in the CHEMLAB-II system (20) was used. We also considered the correlation of the conformational difference with the activity, using a molecular shape analysis method

developed by Hopfinger (23); the molecular conformations of respective compounds were expressed by two values, MSA and CDV. MSA (in ų) was calculated using the van der Waals radii accepted for respective atoms. CDV (in ų) represents the difference van der Waals volume from a reference compound, 1a, for which the common overlap volume was set to the oxazole ring and was calculated using a numerical integration method developed by Pearlstein (24).

Results and Discussion

Molecular Dimensions and Conformations

Although some bond lengths and angles for the same chemical group differ slightly from one to another, the listed values within their estimated standard deviations all seem to lie in the expected region (see Table 3). All the oxazole and benzene rings essentially form planar structures with slight fluctuations, less than 0.02Å. The amide groups also take the usual transconformation ($\Delta\omega_{\rm max}=14.8(3)^{\circ}$ for 2).

Stereoscopic drawings, projected onto the oxazole ring, of a molecular unit for each of the compounds (1-3, 5-12) are presented in Fig. 1. Because both compounds 1 and 5 contain two crystallographically independent molecules per asymmetric unit, the respective molecular conformations are designated a and b. The torsion angles defining the carbamate side chain with respect to the oxazole ring are listed in Table 4. A conformational characteristic observed in 5-phenyl-2-oxazolecarbamate derivatives (1-3, 5, and 8) is the coplanar formation of the oxazole ring with the phenyl and amide planes, ω_1 and ω_2 torsion angles are all in the trans- and cis-regions, respectively. The cis preference of ω_2 is also observed in compounds 6, 7, 9, and 11. This is obviously due to the resonance effect between respective aromatic planes; the average bond lengths of C5-C51 (1.455Å) and C2—N21 (1.368Å) are in the partial doublebonded region. When torsion angles ω_4 and ω_5 are compared

TABLE 4

Conformational torsion angles

	ω1*	ω ₂	ω3	ω4	ω_{5}	ω_6
			deg	rees		
1 a	168.0 (3)	-7.1 (4)	167.2 (2)	-176.8 (2)	88.3 (3)	-94.6 (3)
1b	143.7 (2)	-28.7 (3)	-177.3 (2)	-179.7 (2)	81.1 (3)	-144.8 (3)
2	171.6 (3)	-2.1 (5)	165.2 (3)	-176.0 (3)	87.6 (3)	-92.2 (4)
3	175.0 (2) ^b	-5.6 (4 (174.3 (2)	-175.2 (2)	86.1 (3)	$-96.8\ (3)^{c}$
5 a	171.3 (5)	-12.3 (8)	179.4 (5)	176.2 (5)	-179.1 (S)	-119.0 (6)
5b	-171.5 (5) ^b	4.9 (9)	-176.3 (5)	-177.6 (S)	179.0 (5)	-66.5 (8)°
6		-25.4 (4)	-169.4 (2)	179.5 (2)	-159.6 (2)	-98.5 (3)
7		160.6 (3)	-177.8 (3)	-172.1 (3)	82.5 (4)	-130.0 (4)
8	-160.8 (3) ^b	-4.8 (5)	-178.6 (3)	174.0 (3)	178.2 (3)	100.0 (.)
9	, ,	-1 (1)	-177.5 (6)	-173.8 (6)	172.1 (6)	-157.4 (7)°
10°	174.3 (4) ^b	102.1 (4)	-175.3 (3)	178.8 (3)	89.8 (4)	-97.7 (5)
11°	-174.6 (3) ^b	` ,	176.3 (3)	-176.7 (3)	-109.2 (3)	$-102.4 (4)^{c}$
12°	• • •		-179 (2)	177 (2)	174 (1)	-120 (2)

- "ω₁, 52'-51-5-1; ω₂, 1-2-21-22; ω₃, 2-21-22-24; ω₄, 21-22-24-25; ω₅, 22-24-25-26; ω₆, 24-25-26-27.
- ^b Torsion angles of 52-51-5-1 are listed.
- 'Torsion angles of 24-25-26-27' are listed.

Torsion angles corresponding to those in 1-9 are listed.

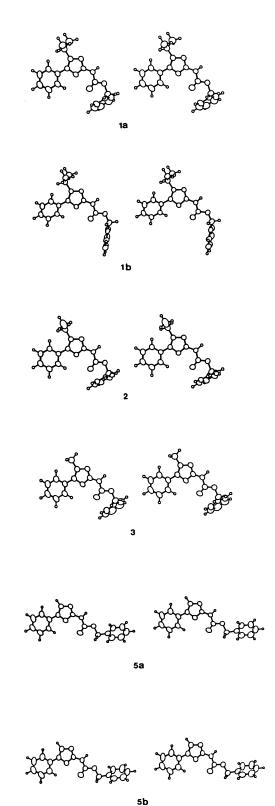


Fig. 1. Stereoscopic projections of 1-3 and 5-12 molecular conformations observed by X-ray analysis, projeted onto the oxazole ring.

with each other, it is also characteristic that ω_4 is restricted to the *trans*-conformation, whereas ω_5 rotates relatively freely. This would result from the difference between the C22-024 and 024-C25 bonding environments. The former bond (average

value, 1.340Å) has a partial double-bond character; therefore, the *trans*-conformation is energetically stable. The latter one (average value 1.457Å) is in the normal, single-bonded region, allowing free rotation about this bond. Possible torsion angles



TABLE 5
Hydrogen bonds and their patterns

	Hydrogen b	onds	Hydrogen bonds Distance Symmetry operation		the drawn handing nation
	Donor at (x, y, z)	Acceptor at	Symmetry operation	(Å)	Hydrogen-bonding pattern
1	N21a	N3b	x, y, z	2.999 (3)	NH N dimer
	N21b	N3a	x, y, z	2.979 (3)	
2	N21	N3	1 - x, -y, 1 - z	2.924 (4)	NH N dimer
3	N21	N3	2 - x, 1 - y, -z	2.909 (3)	NH N dimer
5	N21a	N3a	2 - x, -y, 2 - z	2.808 (7)	NH N dimer
	N21b	N3b	1-x, -y, 2-z	2.807 (7)	
6	N21	N3	-x, 1 $-y$, 1 $-z$	2.838 (3)	NH N dimer
7	N21	01	-x, -y, -z	2.935 (4)	NH O dimer
8	N21	N3	1 - x, -y, 1 - z	2.830 (4)	NH N dimer
9	N21	N3	2-x, $1-y$, $1-z$	2.893 (9)	NH N dimer
10*	N21	N3	x, y, z - 1	3.039 (5)	NH N infinite chain
11*	N21	024	1:5 - x, y - 0.5, 1 - z	2.913 (4)	NH O infinite chain
12*	N21	024	x, 1+y, z	2.97 (2)	NH O infinite chain

Numberings are different from the atomic parameters used for X-ray analyses for the sake of their comparisons with 1-9.

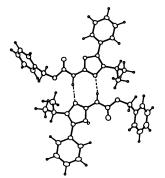


Fig. 2. N-H . . . N hydrogen-bonded dimer formation between compounds 1a and 1b.

of ω_6 could be close to $\pm 90^\circ$ and $\pm 150^\circ$. Among these, the alkyl side chain attached directly to the aromatic ring has preference for the torsion angle of $\pm 90^\circ$. Compounds 1a, 2, 3, 6, 10, and 11 also show this tendency.

Characteristic hydrogen-bonding mode. As summarized in Table 5, four kinds of hydrogen-bonding patterns are formed in the molecular packing of compounds 1-3 and 5-12. Among these, the N12-H ... N9 hydrogen-bonded dimer formation (type A), as exemplified in Fig. 2, is the most probable molecular packing mode for 2-oxazolecarbamate compounds in spite of the differences in crystal packing environments. Only compound 7 shows a different N12-H ... O11 hydrogen-bonded dimer (type B). The predominance of type A over type B implies that the electron-acceptor ability of N3 is stronger than that of the O1 atom. It is interesting to note that the torsion angle ω_2 is largely responsible for the hydrogen bonding modes of types A and B, as shown in the following scheme:

The hydrogen bonding modes observed in compounds 10-12 are basically different from those in 1-9, because of the fundamental differences in their chemical structures.

These hydrogen-bonded dimer formations may also be formed in the solution state, with relatively high concentration. The downfield movements of N12-H proton chemical shifts (δ) were observed in dimethyl sulfoxide- d_6 solutions of 1-9, in proportion to the increase of the concentration (4-10 mM). Their shift values are in the range of 0.04-0.06 ppm, whereas those of 10-12 were not so significant (0.02-0.04 ppm). On the other hand, the temperature dependences of these NH protons at 4 mM concentration showed no hydrogen bonded aggregation, because the $d\delta/dT$ values (4.3 - 7.1 ppb/degree) are too large to consider as a defined conformation. This fact implies that each of 1-9 has the tendency to form a molecular aggregation, via hydrogen bonds, with the increase in the concentration, although they behave like a single molecule in dilute solution.

Conformational analysis of oxazolecarbamate compounds and possible conformation of compound 4. In order to estimate the conformational stabilities of oxazolecarbamate compounds as respective free molecules and, further, to consider the possible conformation of compound 4, it is very important to ascertain whether the conformations observed by X-ray analyses correspond to the energetically stable ones. Contour energy maps for each of the two rotatable torsions, calculated by the molecular mechanics force field method using the atomic coordinates of 5a, are shown in Fig. 3. The filled circles show the conformations observed by X-ray analyses. As seen in Fig. 3a, the low energy region for the (ω_1, ω_2) map has a relatively large rotational freedom. The cis-conformation of ω_2 allows larger freedom than the trans-conformation. The cis preference of ω_2 in the X-ray analyses does not conflict with this result. The X-ray analyses also suggest that free rotation is allowable for ω_5 , whereas ω_4 is constrained to the transconformation. This tendency is ascertained by the energy contour map of (ω_4, ω_5) shown in Fig. 3b. It can be seen from Fig. 3c that there are several minima for the rotational energy of ω_5 and ω_6 , as indicated on the contour map. However, the ω_5 and ω_6 region, close to ($\pm 90^{\circ}$, $\pm 90^{\circ}$), is more stable than that of ω_5 $= 180^{\circ}$ by 1-2 kcal/mol.

The stable torsion angles estimated from the theoretical conformational energy analyses are, on the whole, in good agreement with those observed in the X-ray analyses, suggesting that the molecular conformations revealed by X-ray analyses (Fig. 1) represent the inherent stable forms for the respec-

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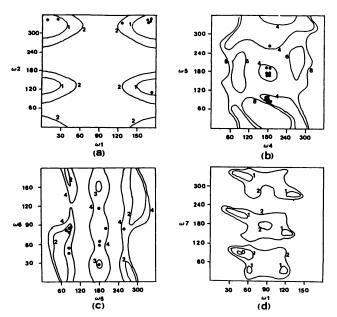


Fig. 3. Contour energy maps of (ω_1, ω_2) (a), (ω_4, ω_5) (b), (ω_5, ω_6) (c), and (ω_1, ω_7) (d). The *number* in the map represents the relative energy value (kcal/mol).

tive compounds and can be used for the analyses of quantitative structure—activity relationships. Encouraged by these results, the most possible conformation of compound 4 proposed is shown in Fig. 4. This conformation is essentially the same as compounds 1a, 2, and 3, except for ω_1 . When 4 takes a conformation at ω_1 of 180°, the strict steric hindrance occurs between the phenyl ring and the *tert*-butyl group, clearly seen in Fig. 3d). In contrast, the conformation at ω_1 of 90° is in the global minimum, allowing rotational freedom for the torsions ω_1 and ω_7 (atoms 3-4-41-42). The most stable conformation is at ω_1 of 45° and ω_7 of 90°, as shown in Fig 4.

Structure-Activity Relationships

For the purpose of geometric analysis for the inhibitory activity, it is important to consider which groups participate in the binding to aldose reductase. Although the absence of detailed structural information on the enzyme frustrates this consideration at present, the following discussion on the structural and conformational comparisons with the inhibitory activity leads to the conclusion that the benzyl 5-phenyl-2-oxazolecarbamate skeleton and its spatial conformation are absolutely necessary for the emergence of activity.

The replacement of the 5-phenyl ring (1-5) with the methyl (7 and 9) or tert-butyl groups (6) causes drastic decrease in the inhibitory activity, implying the importance of the hydrophobic aromatic ring at the oxazole C5-site and its coplanar formation with the oxazole ring (compare 1-3 with 4). As exemplified by the compounds 10 through 12, the structural modification of the oxazole ring itself (12) or the binding site of the phenyl and carbamate chains (10 and 11) also causes drastic three-dimensional changes (see Fig. 1), consequently leading to complete inactivation. On the other hand, the existence of the benzyl end-group and its spatial orientation with respect to the oxazole ring are also important for activation (compare compounds 1-3 with 5 and 8). It is conceivable from these facts that the binding site of aldose reductase for 2-oxazolecarbamate compounds has a well defined spatial orien-

tation (binding specificity) and that the hydrophobic interactions between the phenyl and the benzyl rings, as well as the hydrogen bonds in which the N and/or O atoms of oxazole ring and amide group participate, are essential for the best binding with the receptor.

It is of special importance to note that the highly active benzyl 5-phenyl-2-oxazolecarbamates (1-3) all show the following conformational similarities, irrespective of the substituents at the oxazole C4-site: $\omega_1 = \sim 180^\circ$, $\omega_2 = \sim 0^\circ$, $\omega_3 = \omega_4 =$ ~180°, $\omega_5 = \sim 90$ °, and $\omega_6 = \sim -90$ °. This conformation is also energetically stable, as seen from Fig. 3. Therefore, it is reasonable to conceive that 1-3 represent the best-fit forms for the binding pocket of the reductase (the surface contour mapping for 1a is shown in Fig. 5). In addition to the conformational similarities, compounds 1-3 show similar characteristics in their dipole moments; their dimensions and directions with respect to the oxazole ring fixed at the orientation shown in Fig. 1 are in the narrow range of 1.2-1.5 D for the x (horizontal, μ_x), -0.3-0.1 D for the y (vertical, μ_y), and 1.3-1.7 D for the z direction (perpendicular to x and y, μ_z). The other compounds (4-12) exhibited dipole moments more or less deviated from this range. On the other hand, the inhibitory activity of benzyl 5-phenyl-2-oxazolecarbamate analogues (1-5) appears to be related to the bulkiness of the alkyl group attached at the oxazole C4-site, in addition to the molecular conformations; the activity increases in the order of -H (5) < $-C(CH_3)_3$ (4) < —CH₃ (3) < —CH₂CH₃ (2) < —CH(CH₃)₂ (1) (their Taft steric substituent constants (25) are -1.12, 1.43, 0.0, 0.08, and 0.48, respectively). A reason for compound 4 having less activity than 1 is due to the deviation of the 2-phenyl ring from the coplanar formation with the oxazole ring.

In order to clarify the relationship between the above-mentioned chemical and conformational characteristics and their activities, a QSAR analysis was carried out in which compounds 1-9 were considered. The results are given in Table 6. The contribution of $\log P$ (lipophilicity) to the activity is relatively poor when all compounds are taken into account (compare Eq.

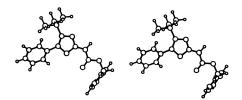


Fig. 4. Most probable molecular conformation of compound 4 proposed from theoretical energy calculations.

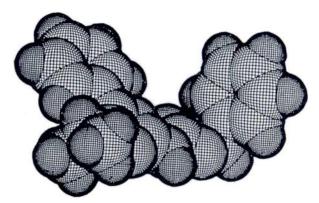


Fig. 5. Surface-contoured molecular conformation of compound 1a.

TABLE 6
Correlation coefficients of QSAR parameters and correlation equations for inhibitory activities

	log P	E.40	E,56	μ_x	μγ	μ_z	MSA	CDV	
Linear correlation	on coefficients use	d for QSAR and	alyses						
rc	0.516	0.512	0.684	0.489	0.326	0.842	0.696	0.913	
						n⁴	r	SD*	
Correlation equa	ations for inhibitor	y activities							
Eq. 1: log(1/l0	C_{50}) = $-0.0417 - 10^{-2}$	0.0327 CDV				11	0.913	0.457	
	(0.2352)								
Eq. 2: log(1/l	C_{50}) = $-0.0424 +$	•				11	0.913	0.457	
		(0.1801)							
Eq. 3: log(1/l	C_{50}) = $-0.9091 +$					11	0.915	0.451	
	(1.7530) ((0.0065)						
Eq. 4: log(1/l	C_{50}) = -0.2808 -					11	0.967	0.286	
		(0.0988) (0.				4.4			
Eq. 5: log(1/10	C_{50}) = $-0.7196 + 0.0000$	-				11	0.931	0.408	
E - 0 1 14 11		(0.1734) (0		05 001		44	0.070	0.040	
Eq. 6: log(1/10	C_{50}) = $-0.3324 + 0.0000$					11	0.976	0.243	
	(0.3022)	U.1443) (C).1174) (0.00	39)					

^{*} Taft's steric substituent constant at oxazole C4-site.

1 with Eq. 2). A good correlation is obtained with the CDV (Eq. 1) rather than with the MSA (Eq. 3), implying that the inhibitory activity is largely dependent on the molecular conformation. This conformational importance for the activity emergence can also be suggested from the correlation with the dipole orientation of the z direction (μ_z) (Eq. 4). Furthermore, the existence of the phenyl ring at the oxazole C5-site is also important for the effective activity emergence (see Eqs. 5 and 6).

The present results derived from the combination of X-ray crystallographic and QSAR analyses clearly showed that the molecular conformation is very important for the inhibitory activity of oxazole compound. Therefore, it is very interesting to consider how this insight relates to the understanding of another potent aldose reductase inhibitor. Kador et al. (16, 26) have proposed a possible interaction mode between aldose reductase and its inhibitor, based on the electrostatic and steric features of various inhibitors. According to this model, the binding pocket of the enzyme consists of two hydrophobic (H1 and H2) and a charge-transfer (CT) site, and the relative disposition among these three sites is very important for the tight binding with the inhibitor. The CT site, which is located adjacent to the two hydrophobic binding sites, is responsible for the hydrogen bonding with the polar atoms within the inhibitor. It is interesting to see that the conformational significance concluded from the present study can be well supported by this binding model. Judging from the comparison of the molecular conformation shown in Fig. 5 with the schematic diagram of the binding model (see Fig. 3 of Ref. 16), the coplanar 5-phenyloxazole moiety (A), terminal benzyl group (B), and the carbamate group (C) appear to bind well with the H1, H2, and CT sites of the enzyme, respectively. The distances of A(center)-B(center), A(center)-CT(C22), and CT(C22)-B(center) are 7.88, 3.12, and 4.30 Å, respectively, and are in the region proposed for other potent inhibitors. Furthermore, the present molecular shape analyses have proposed that the H1 site is at almost right angles to the H2 one (the averaged dihedral angle between A and B planes is 103.3° for 1-3).

Drug design of more potent analogues is now in progress, based on the present insights.

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^b Taft's steric substituent constant at oxazole C5-site.

^c Correlation coefficient with inhibitory activity.

d Number of data used in regression.

Standard deviation from regression line.

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